

Retmańska K., Pobereżny J., Wszelaczyńska E., Gościnna K., Ropińska P. 2023.

Organoleptic characteristics and the total glycoalkaloid content
of edible potato tubers depending on a cultivation technology and storage.

J. Elem., 28(1): 7-25. DOI: 10.5601/jelem.2022.27.4.2359



RECEIVED: 28 November 2022 ACCEPTED: 10 January 2023

ORIGINAL PAPER

ORGANOLEPTIC CHARACTERISTICS AND THE TOTAL GLYCOALKALOID CONTENT OF EDIBLE POTATO TUBERS DEPENDING ON A CULTIVATION TECHNOLOGY AND STORAGE*

Katarzyna Retmańska¹, Jarosław Pobereżny¹, Elżbieta Wszelaczyńska¹, Katarzyna Gościnna¹, Paulina Ropińska²

¹ Institute of Agri-Foodstuff Commodity
 ² Department of Agricultural Chemistry
 Bydgoszcz University of Science and Technology, Poland

Abstract

Criteria for the selection of edible potato varieties are such characteristics as the skin and flesh color, palatability and the tendency of the flesh to darken, which are subjectively evaluated by the consumer, as well as the content of undesirable substances like solanine. The quality of the edible potato depends primarily on the environmental conditions and cultivation technology. Magnesium has a considerable influence on the size and quality of the potato tuber yield. If potato plants are grown under stress conditions, a biostimulant should be used. The purpose of the research was to determine the influence of magnesium, application of biostimulant and storage time on organoleptic properties of potato tubers of the edible variety called Gala as well as the total content of glycoalkaloids (TGA) in tubers. The experimental factors were: assessment date (immediately after harvest as well as after 6 months of storage in 4°C and RH 95%), magnesium fertilization (0, 30, 60, 90 kg ha⁻¹), biostimulator application (0, 1.5, 3.0 l ha⁻¹). The study showed that the application of magnesium during the potato growing season caused a change in the intensity of tuber flesh color to a more yellow color and increased the content of TGA. The greatest effect in this regard was obtained with the dose of 90 kg MgO ha-1. In addition, the application of magnesium reduced the darkening processes of both raw and cooked tubers. The best results in this regard were obtained after the application of magnesium in soil at 30 and 60 kg ha⁻¹. The application of the biostimulant had a positive effect on the distinguishing features of the consumption value: taste and smell. The best taste and neutral

Katarzyna Retmańska, M.Sc., Institute of Agri-Foodstuff Commodity, Bydgoszcz University of Science and Technology, Kaliskiego 7 Bydgoszcz, Poland, e-mail: katarzyna.retmanska @pbs.edu.pl

^{*} Research supported by the Ministry of Science and Higher Education of Poland as part of the statutory activities (BS-48/2014) of the Department Microbiology of Food Technology, Bydgoszcz University of Science and Technology.

smell were obtained by potato tubers from objects where the biostimulant was applied at a dose of 1.5 l ha⁻¹. Application of the biostimulant increased the TGA content in tubers. Long-term storage generally deteriorated the quality of the Gala potato variety tubers in terms of all the tested organoleptic characteristics and increased the TGA content.

Keywords: edible potato, consumption value, oxidative potential, magnesium, biostimulant, storage, solanine

INTRODUCTION

Potato, along with wheat, rice and corn, is ranked among the most important crops. Owing to the high nutritional value of potato tubers, the plant continues to be a dietary staple food for many people (Devaux et al. 2014, Alamar et al. 2017). Consumers seek on the market varieties with good palatability. Characteristics such as the flesh and skin color and the tendency of the flesh to darken are also criteria for their selection. However, it should be remembered that the sensory properties of tubers are assessed by consumers subjectively (Wszelaczyńska et al. 2017).

Potato quality is influenced by the environmental conditions and cultivation technology. No less important factors are genetic predispositions, as well as duration and conditions of storage (Rytel et al. 2008, Zgórska, Grudzińska 2012). Among the elements of great importance for the size and quality of the potato tuber yield there is the element magnesium. Its absence in the plant causes the inhibition of the plant's growth and development, and negatively affects the culinary characteristics of the tubers (Osowski et al. 2017). The potato quality is also influenced by the use of biostimulants in its cultivation. Biostimulants are responsible for the activation and control of many vital processes. They also affect the nitrogen use by crop plants (Sawicka et al. 2016).

There are specific requirements for potatoes intended for direct consumption. Potato tubers of edible varieties are evaluated in terms of morphological, physiological and organoleptic characteristics (Leszczyński 2000, Lisińska 2006, Zgórska, Grudzińska 2012). Table potatoes should be characterized by shapely tubers with smooth skin and only slightly darkening flesh (on a scale of 9: raw flesh >6.5 and after cooking >7.5) – Zgórska, Grudzińska (2012), Wszelaczyńska et al. (2017). They should also be characterized by good taste and neutral smell. The high quality of potatoes is also determined by the nutritional value of the tubers and the low content of undesirable components (Lisińska 2006, Zarzecka, Gugala 2011, Wszelaczyńska et al. 2013). The main components affecting the safety and taste of potato tubers are glycoalkaloids (α - and β -solanine, chaconin and leptin) – Ginzberg et al. (2009), Haase (2010), Rytel (2012). Their accumulation in tubers is determined by the genetic composition of a potato plant, the climate and soil conditions, cultivation technology and even the post-harvest treatment of tubers

(Şengül et al. 2004, Machado et al. 2007, Tajner-Czopek et al. 2008, Wszelaczyńska et al. 2020). Due to their high toxicity, the U.S. Food and Drug Administration (FDA) has set the maximum permissible total content of glycoalkaloids in potatoes at 200 mg kg⁻¹ fresh weight (FW) and the intake level at 1 mg kg⁻¹ body weight. It is also important to note that the TGA content in potatoes above 100 mg kg⁻¹ fresh weight contributes to taste deterioration (Ginzberg et al. 2009, Wszelaczyńska et al. 2020).

Edible potatoes are stored for up to 9 months, so storage conditions are important for maintaining good tuber quality. Consumable varieties are recommended to be stored at a constant temp. of 4°C and 95% RH. Not maintaining the recommended temperature, humidity or inadequate light access can promote excessive accumulation of TGAs in tubers (Chung et al. 2004, Rezaee et al. 2011, Wszelaczyńska et al. 2020).

The purpose of the study was to determine the effect of magnesium fertilization, application of a biostimulant and storage time on the TGA content and organoleptic characteristics of the edible potato variety Gala.

MATERIAL AND METHODS

The research material was the early variety of edible potato "Gala". A three-factor field experiment (2015-2017) was established using the randomized sub-block design at the Agricultural Research Station in Mochelek (Kuyavian-Pomeranian Voivodeship) belonging to the Bydgoszcz University of Science and Technology (53°13′N, 17°51′E).

The experimental factors were:

- assessment date (A) immediately after harvest and after 6 months of storage;
- magnesium fertilization (B) MgO doses: 30, 60, 90 kg ha⁻¹;
- biostimulant application (C) −1.5, 3.0 l ha⁻¹.

Mineral magnesium was applied to the soil before potato planting in the form of 16% magnesium sulfate (MgSO₄) with constant NPK fertilization at doses: 100 kg N ha⁻¹ (ammonium nitrate) - 34%; 120 kg K₂O ha⁻¹ (potassium sulfate) - 50%; 80 kg P₂O₅ ha⁻¹ (triple superphosphate) - 46%.

The biostimulant Protifert LMW was applied three times: to plants with a height of 15 cm, immediately after flowering and 2 weeks after flowering. Biostimulant Protifert LMW is a liquid fertilizer containing a peptide of organic origin and amino acids. It is produced from natural raw materials. It is characterized by systemic translocation in the plant and rapid absorption due to its water solubility. Protifert LMW stimulates plant absorption of nutrients, increases the effectiveness of plant protection products and improves soil structure.

In each year of the field experiment, potatoes were planted with a mechanical planter in the last ten days of April. The experiment was established in plots of 33.6 m² with a row spacing of 0.75 x 0.35 m. 22.4 m² were assigned for harvesting. In each year, the preceding crop for the potato was cereal. In the autumn, manure was applied at a dose of 25 Mg ha⁻¹.

Chemical protection of plants against pests and diseases as well as agrotechnical treatments were carried out in accordance with the principles of optimal agrotechnology for potatoes. During the cultivation, the following potato protection agents were applied: herbicide – Afalon 50WP (linuron) in a dose of 2 l ha⁻¹, fungicides – Ridomil Gold MZ 67.8 WG (metalaxyl-M + mancozeb) dosed at 2 kg ha⁻¹ and Acrobat MZ 69 WG (dimetomorf + mancozeb) dosed at 2 kg ha⁻¹ as well as insecticides – Mospilan 20 SP (acetamiprid) in a dose of 80 g ha⁻¹ and Calypso 480SC (tiachloprid) in a dose of 0.08 l ha⁻¹.

The tubers were harvested at full maturity. Samples of 10 kg of tubers were taken from each experimental site for analytical tests immediately after harvesting and after long-term storage. The samples were placed in a controlled-atmosphere storage facility for a period of 6 months (October-March). A constant temp, of 4°C and a relative humidity of 95% were maintained throughout the storage period. The tubers were stored in two chambers (Thermolux Chłodnictwo Klimatyzacja, Raszyn, Polska) in the Institute of Food Commodities of Bydgoszcz University of Science and Technology. Each experimental storage chamber was 2 m high, 2 m wide and 3.8 m long. The tank shell material was milky-white translucent polypropylene sheets. Milk-white translucent polypropylene sheets as the tank shell material ensure such benefits as flame retardancy, moisture and light resistance, as well as providing heat isolation. In addition, in order to simulate the temperature environment and decrease heat loss, the experimental storage chamber was coated with 20 mm thick foam insulation material. The chambers were equipped with an automatic system for maintaining humidity in the storage chambers. When it fell below 95%, the system switched on and fogged the air through nozzles. The experiment was set up on a Luvisol (LV) soil, which had a fine sandy loam texture. The soil parameters are shown in Table 1. Average monthly air temperature and total rainfall in the years 2015-2017 and in the multi-year period 1996-2014 are shown in Table 2.

Throughout the 2015 growing season, with the exception of August and September, the temperature was below that for the multi-year period average. There were also small amounts of precipitation compared to the multi-year total. During the 2016 and 2017 growing seasons, air temperatures were similar to those for the multi-year period. However, during the 2016 growing season, May, August and September received less precipitation than for the multi-year period. In contrast, less precipitation in 2017occurred only in May and August.

Consumer evaluation of potato tubers was performed immediately after harvest and after 6 months of storage. The evaluation was carried out

 ${\it Table 1}$ Soil parameters before the field experiment (average of 2015-2017)

| Parameter | Amount | Abundance |
|--|--------|-----------------|
| pH _{H2O} | 6.6 | slightly acidic |
| pH _{KCl} | 6.1 | slightly acidic |
| Organic carbon (g kg ⁻¹) | 8.75 | - |
| Total nitrogen (g kg ⁻¹) | 0.78 | - |
| Absorbable forms of phosphor (mg kg ⁻¹) | 27.0 | poor |
| Absorbable forms of potassium (mg kg ⁻¹) | 49.0 | very poor |
| Absorbable forms of magnesium (mg kg ⁻¹) | 23.0 | very poor |

Table 2 Average monthly air temperature and total rainfall in the years 2015-2017 and in the multi-year period 1996-2014

| | Air temperature (C°) | | | Rainfall (mm) | | | | |
|-----------|----------------------|------|------|---------------|------|-------|-------|-----------|
| Month | 2015 | 2016 | 2017 | 1996-2014 | 2015 | 2016 | 2017 | 1996-2014 |
| April | 7.5 | 8.3 | 6.8 | 8.1 | 15.6 | 28.7 | 40.8 | 28.7 |
| May | 12.4 | 14.7 | 13.4 | 13.2 | 21.6 | 51.4 | 56.3 | 61.1 |
| June | 15.6 | 17.7 | 16.8 | 16.3 | 33.0 | 98.1 | 54.3 | 53.1 |
| July | 18.5 | 18.3 | 17.7 | 18.7 | 50.4 | 133.8 | 118.9 | 87.1 |
| August | 20.9 | 16.4 | 14.3 | 17.8 | 20.3 | 55.3 | 19.4 | 67.0 |
| September | 13.8 | 14.3 | 13.0 | 13.0 | 52.4 | 19.4 | 78.4 | 66.5 |
| Average | 14,8 | 15.0 | 13,7 | 14.5 | 32.2 | 64.5 | 61.4 | 60.6 |

by a permanent team of 10 people in accordance with PN-ISO 8586 (Roztropowicz et al. 1999).

The scope of organoleptic evaluation included:

- smell (scale of 4 points: 1 neutral, 4 foreign) according to Zimnoch-Guzowska, Flis (2006);
- taste (scale of 9 points: 9 very good, 1 bad) according to Wroniak (2006);
- color (scale of 6 points: 1 white flesh, 6 points dark yellow) according to Keutgen et al. (2014);
- darkening of tubers after cooking: after 10 min and 24 h (scale of 9 points: 9 – non-darkening, 1 – black) according to Zgórska (2013);
- darkening of raw tubers after 10 min and 4 h (scale of 9 points:
 9 non-darkening, 1 black) according to Zgórska (2013).

To determine the TGA content, the potato material was extracted according to the Bergers (1980) method, in which the lyophilized material is dissolved in 0.5 mL of 7% (v/v) phosphoric acid and kept at -20°C until testing. To determine the total TGA content, 200 μ L of the extract was added to 1 mL of 0.03% (w/v) paraformal dehyde in concentrated phosphoric acid. After color unfolding (20 min), the absorbance was tested (600 nm on Shimadzu UV-1800 Japan) and the TGA concentration was measured on the basis of α -solanine and α -chaconine (Sigma-Aldrich, St. Louis, MO, USA) standard curves.

The test results were subjected to statistical calculations. The method of variance analysis for three-factor experiments was used for statistical calculations. The Tukey's test was used to assess the significance of differences, at the level of $\alpha=0.05$. The calculations were made in the FR-ANALWAR program based on Microsoft Excel. In order to obtain a synthetic picture of the overlapping dependencies between the examined features, a simple linear correlation (Pearson) analysis was performed.

RESULTS AND DISCUSSION

Smell

The results of the post-cooking smell of the Gala variety potato tubers depending on the cultivation technology and the date of evaluation are presented in Table 3. As can be seen from the data, regardless of the factors used, the tubers of the tested variety were characterized by a neutral smell and received an average score of 1.2 points. Polish potato varieties are generally characterized by a neutral smell (Zimnoch-Guzowska, Flis 2006, Zarzecka et al. 2010, Zarzecka, Gugała 2011). Many authors (Zarzyńska, Goliszewski 2006, Styszko, Kamasa 2007, Zarzyńska, Wroniak 2007, Pyryt, Kolenda 2009) state that potato tuber smell is a stable trait, but can be affected by genetic conditions. On the other hand, according to Ciećko et al. (2005) the smell of tubers after cooking depends mainly on the esters, alcohols, aldehydes, pyrazines and volatile sulfur compounds contained in them.

In the conducted study, magnesium fertilization and application of a biostimulant during the growing season significantly affected the tuber flesh smell both immediately after harvest and after storage. Each of the applied doses of magnesium caused the tuber flesh smell to change to a more noticeable one compared to the control object. The most noticeable smell was obtained after the application of a dose of 60 kg MgO ha⁻¹ both after harvest and after storage. The opposite view is expressed by Rogozińska et al. (2004) in their study, in which increasing doses of magnesium fertilization increased the concentration of this element in the tubers, which contributes to improving their smell. On the other hand, application of the biostimulant

 ${\it Table 3}$ Sensory evaluation of cooked potato tubers of the Gala variety depending on the cultivation technology and evaluation date (average of 2015-2017)

| MgO fertilization doses (B) (kg ha¹) | | | Pota | to tuber eva | luation dat | e (A) | |
|--|------------|--------------------------|---------------|----------------|---------------------|-------|-------|
| | | direc | tly after ha | rvest | after storage | | |
| | | smell | taste | color | smell | taste | color |
| | | Without bio | stimulant ap | plication, con | trol (C) | | |
| | 0 | 1.0 | 7.0 | 4.6 | 1.0 | 6.9 | 4.4 |
| : | 30 | 1.4 | 7.0 | 4.7 | 1.4 | 8.0 | 4.5 |
| (| 30 | 2.0 | 8.0 | 5.1 | 2.0 | 7.4 | 4.8 |
| ę | 90 | 1.4 | 7.4 | 5.0 | 1.4 | 7.1 | 4.8 |
| Ave | erage | 1.4 | 7.3 | 4.8 | 1.4 | 7.3 | 4.6 |
| | | Biostimul | ant applicati | on – 1.5 l ha | 1 ⁻¹ (C) | | |
| | 0 | 1.0 | 7.0 | 4.5 | 1.0 | 7.3 | 4.5 |
| 3 | 30 | 1.0 | 7.5 | 4.5 | 1.0 | 7.5 | 4.5 |
| | 30 | 1.0 | 8.3 | 5.0 | 1.0 | 8.0 | 4.8 |
| | 90 | 1.0 | 7.3 | 5.0 | 1.0 | 8.3 | 5.0 |
| Ave | erage | 1.0 | 7.5 | 4.8 | 1.0 | 7.8 | 4.7 |
| | | Biostimula | ant applicati | on – 3.0 l ha | 1 ⁻¹ (C) | | |
| | 0 | 1.0 | 7.8 | 4.3 | 1.0 | 7.0 | 4.5 |
| (| 30 | 1.0 | 6.5 | 4.8 | 1.0 | 8.0 | 4.5 |
| (| 30 | 1.0 | 7.0 | 5.0 | 1.0 | 7.3 | 4.6 |
| Ę | 90 | 1.0 | 7.5 | 5.3 | 1.3 | 7.3 | 4.8 |
| Ave | erage | 1.0 | 7.2 | 4.8 | 1.1 | 7.4 | 4.6 |
| | | | Averag | ge | | | |
| | 0 | 1.0 | 7.3 | 4.4 | 1.0 | 7.0 | 4.5 |
| 9 | 30 | 1.1 | 7.0 | 4.7 | 1.1 | 7.8 | 4.5 |
| (| 30 | 1.3 | 7.8 | 5.0 | 1.3 | 7.5 | 4.7 |
| (| 90 | 1.1 | 7.4 | 5.1 | 1.2 | 7.6 | 4.8 |
| Ave | erage | 1.2 | 7.3 | 4.8 | 1.2 | 7.5 | 4.6 |
| NIR _{0.05} (Tukey Smell | 's test) | | | | | | |
| A – ns | B – 0.16 | C - 0.13 | | | | | |
| A - ns B/A - ns | A/B - ns | C = 0.13 B/C = 0.27 | | | | | |
| A/C - ns | C/B - 0.27 | B/C = 0.27 B/C = 0.27 | | | | | |
| Taste | C/D = 0.27 | DIC - 0.21 | | | | | |
| A – ns | B - 0.39 | C - 0.13 | | | | | |
| A - Hs B/A - 0.55 | A/B - 0.54 | B/C – ns | | | | | |
| A/C - ns | C/B - 0.26 | B/C - 0.44 | | | | | |
| Color | C/D 0.20 | DIO 0.44 | | | | | |
| A – ns | B - 0.19 | C-ns | | | | | |
| A - ns B/A - ns | A/B - ns | C - Hs B/C - 0.13 | | | | | |
| A/C - 0.20 | C/B - 0.18 | B/C = 0.13 B/C = 0.24 | | | | | |
| ns = non-signifi | | 5/0 0.24 | | | | | |

ns-non-significant

in any amount contributed to the improvement of smell. The results of organoleptic evaluations indicate that the smell of cooked tubers did not change after long-term storage. In contrast, according to Krochmal-Marczak et al. (2016), long-term (8 months) storage worsened cooked tuber flesh smell.

Taste

The results for taste scoring are shown in Table 3. Of all the characteristics included in consumer evaluation, taste and aroma impressions are the most subjective. Wszelaczyńska et al. (2013) state that organoleptic evaluation should include palatability, which is defined as the set of taste and smell sensations experienced during consumption. Zarzecka et al. (2010) and Zarzecka, Gugała (2011) report that Polish varieties are mostly characterized by very good palatability. Our results confirm this as the studied variety was characterized by good palatability. The analysis of variance showed that the cultivation technology had a significant effect on the taste of the of the Gala variety tubers tested after cooking. Both the application of different magnesium fertilization and biostimulant doses improved the taste of the tubers after cooking. Increasing magnesium fertilization resulted in some improvement in the taste of cooked tubers compared to the control, and the best result in this regard was achieved by the magnesium dose of 60 kg MgO ha⁻¹. Flis et al. (2012) report that taste is influenced by the minerals contained in the tubers. Application of the biostimulant at 1.5 l ha⁻¹ significantly improved tuber flavor compared to the control. Increasing the application dose to 3 l ha⁻¹ had no effect on the taste of the tested variety. A similar opinion is held by Płaza (2010), who claims that the palatability of potato tubers changes significantly under the influence of environmental conditions. Additionally, Grudzińska et al. (2016) proved significant influence of the cultivation technology on taste as an organoleptic evaluation discriminator. Our study did not confirm a significant effect of TGA on potato flavor, as there was no significant correlation between the TGA content and flavor (Tables 4 and 5). However, it should be noted that the content of TGA in our study was at a low level and many authors (Ginzberg et al. 2009, Wszelaczyńska et al. 2020) suggest the taste deteriorates after exceeding 100 mg kg⁻¹. Long-term storage did not change the taste of tubers of the studied variety. Krochmal-Marczak et al. (2016) reported opposite findings in their study, where eight-month storage caused deterioration of the taste of the three edible varieties tested: Americana, Denar, and Lord.

Color

It is known that the color of potato flesh is a varietal feature. It can be white, creamy or yellow with a range of intermediate shades. Its determination serves primarily to satisfy consumer tastes (Gugała, Zarzecka 2007, Rytel et al. 2008, Pyryt, Kolenda 2009, Płaza 2010, Zarzecka et al. 2010,

 ${\it Table \ 4}$ Linear correlation analysis (Pearson) of the studied characteristics of the Gala potato variety directly after harvest

| Specification | Smell | Taste | Color | DR 10 min | DR 4 h | DC 10 min | DC 24 h |
|---------------|--------|-------|-------|--------------|--------|--------------|---------|
| Taste | 0.384 | | | | | | |
| Color | 0.428 | 0,517 | | | | | |
| DR 10 min | ns | ns | 0.356 | | | | |
| DR 4 h | ns | ns | 0.370 | ns | | | |
| DC 10 min | ns | ns | ns | ns | ns | | |
| DC 24 h | -0.460 | ns | ns | ns | ns | 0,646 | |
| TGA | 0.390 | ns | 0.422 | ns | ns | ns | -0,336 |

Indicates that the correlation is significant at the 0.05 probability level >0.334. ns – non-significant, DR 10 min – darkening of raw potatoes after 10 min, DR 4 h – darkening of raw potatoes after 4 h, DC 10 min – darkening of cooked potatoes after 10 min, DC 24 h – darkening of cooked potatoes after 24 h, TGA – total glycoalkaloids content

| Specification | Smell | Taste | Color | DR 10 min | DR 4 h | DC 10 min | DC 24 h |
|---------------|--------|-------|-------|--------------|--------|--------------|---------|
| Taste | ns | | | | | | |
| Color | ns | ns | | | | | |
| DR 10 min | -0.430 | ns | ns | | | | |
| DR 4 h | ns | ns | ns | ns | | | |
| DC 10 min | ns | ns | ns | ns | ns | | |
| DC 24 h | ns | ns | ns | ns | ns | 0.512 | |
| TGA | ns | ns | 0.433 | ns | ns | -0.399 | -0.430 |

indicates that the correlation is significant at the 0.05 probability level >0.334. ns – non-significant, DR 10 min – darkening of raw potatoes after 10 min, DR 4 h – darkening of raw potatoes after 4 h, DC 10 min – darkening of cooked potatoes after 10 min, DC 24 h – darkening of cooked potatoes after 24 h, TGA – total glycoalkaloids content

Zarzecka, Gugala 2011). This feature is therefore a critical physical property for commercial classification and purchasing decisions. The color parameter is also the primary function of quantifying freshness, which it is particularly important for raw potatoes intended for direct consumption (Kidoń, Czapski 2010, Wszelaczyńska et al. 2013, Keutgen et al. 2014). Edible varieties of potatoes registered in Poland are distinguished by yellow and light yellow color (Zarzecka, Gugała 2011, Wszelaczyńska et al. 2013), which is confirmed by the conducted research. It was shown that the agrotechnical factor applied in the experiment, in the form of magnesium fertilization, signifi-

cantly modified the flesh color of the studied tubers (Table 3). Gugała, Zarzecka (2007) report that the application of various factors during the growing season can cause a change in the flesh color. On the other hand, according to Hamouz et al. (2006), the flesh color depends on the organic acid content and, according to Zimnoch-Guzowska and Flis (2006) and Tilahun et al. (2020), on the concentration of carotenoids, anthocyanins and solanine. Some authors (Tilahun et al. 2020) showed a significant correlation between the raw tuber's flesh color and tuber's solanine content. In our research, immediately after harvest and following long-term storage, correlation coefficients also indicate a significant relationship between the color and TGA content, at r = 0.422 and r = 0.433, respectively (Tables 4 and 5). In the study, each of the applied doses of magnesium during the growing season resulted in a change in the hue of the flesh color of the Gala variety to more yellow than in the control. Similar results were obtained by Rogozińska et al. (2004) in an experiment with the edible variety Mila. The authors showed that the application of magnesium to plants during the growing season included a change in the intensity of flesh coloration of the studied variety. No significant effect of long-term storage on tuber color change was observed. A different opinion is declared by Płaza et al. (2010), Krzysztofik and Skonieczny (2010), Pobereżny and Wszelaczyńska (2011), Wszelaczyńska and Pobereżny (2011), Wszelaczyńska et al. (2013), who maintain that long-term potato storage contributes to changes in the intensity of potato tuber flesh coloration.

Tuber darkening

The results concerning the propensity of the flesh of the Gala variety raw tubers to darken evaluated both 10 min and 4 h after cutting are shown in Table 6. It should be emphasized that the studied variety is genetically characterized by low propensity to darkening (Ciećko et al. 2005, Zgórska et al. 2006). Regardless of the cultivation technology used, immediately after harvest, the propensity of raw flesh to darken after 10 min was assessed at 8.7 points, and after 4 h – at 8.0 points. The applied magnesium fertilization and spraying with the biostimulant during the growing season significantly reduced the tendency for flesh to darken after 10 minutes. In this regard, doses of 60 kg MgO ha and 1.5 l ha were the most effective, respectively. No such results were obtained regarding the evaluation of raw tubers after 4 hours. A similar view is expressed by Wszelaczyńska (2004), Zgórska et al. (2006) and Keutgen et al. (2014), who claim that the propensity of the flesh of raw tubers to darken depends mainly on agrotechnical factors. Of a different opinion are Gugała and Zarzecka (2007), who proved that the method of potato cultivation and care had no effect on the darkening of raw tuber flesh. In addition, our study showed a significant but low correlation (P<0.05) between the darkening process of raw tubers at 10 min and 4 h after cutting and color: r = 0.356 and 0.370, respectively (Table 4). This relationship indicates that flesh color is affected by the course of non-enzymatic

Table 6 Evaluation of darkening after 10 min and 4 h of raw potato tubers of the Gala variety depending on the cultivation technology and evaluation date (average of 2015-2017)

| directly after 10 min after 2 min after 3 min after 4 min a | MgO | | I | Potato tuber eva | aluation date (A |) | | | | |
|--|--------------------------|---|------------------|--------------------|----------------------|-----------|--|--|--|--|
| Without biostimulant application, control (C) | | | directly af | ter harvest | after s | torage | | | | |
| 0 8.8 7.9 8.8 7.2 30 8.6 8.1 8.6 7.7 60 8.9 8.4 8.8 7.6 90 8.6 8.1 8.7 7.7 Average 8.7 8.1 8.7 7.6 Biostimulant application − 1.5 1 ha⁻¹ (C) 0 8.9 7.9 8.8 7.2 60 8.9 8.0 8.9 7.2 60 8.9 8.0 8.9 7.2 60 8.9 8.0 8.9 7.2 60 8.9 8.0 8.0 8.9 7.2 Average 8.8 8.0 8.6 7.2 Average 8.8 8.0 8.6 7.2 Average 8.8 8.0 8.8 7.4 Biostimulant application − 3.0 1 ha⁻¹ (C) 0 8.6 8.2 8.4 7.2 30 8.8 7.8 8.3 7.3 60 8.9 7.9 8.6 7.6 90 8.5 8.0 8.2 7.6 Average 8.7 8.0 8.2 7.6 4 90 8.5 8.0 8.2 7.6 Average 8.7 8.0 8.2 7.6 Average 8.8 8.0 8.7 7.4 Significant of the state of the | (kg | g ha ⁻¹) | after 10 min | after 4 h | after 10 min | after 4 h | | | | |
| Section Sec | | Without biostimulant application, control (C) | | | | | | | | |
| 60 8.9 8.4 8.8 7.6 90 8.6 8.1 8.7 7.7 Biostimulant application − 1.5 l ha¹ (C) 0 8.9 7.9 8.8 7.7 30 8.5 8.0 8.9 7.2 60 8.9 8.0 9.0 7.5 90 8.8 8.0 8.6 7.2 Average 8.8 8.0 8.8 7.4 Biostimulant application − 3.0 l ha¹ (C) 0 8.6 8.2 8.4 7.2 30 8.6 8.2 8.4 7.2 30 8.8 7.8 8.3 7.3 60 8.9 7.9 8.6 7.6 90 8.5 8.0 8.2 7.6 Average 8.7 8.0 8.4 7.4 30 8.6 8.0 8.7 7.4 30 8.6 8.0 8.7 7.4 30 8.6 8.0 8.6 7.5 Average 8 | | 0 | 8.8 | 7.9 | 8.8 | 7.2 | | | | |
| Section | | 30 | 8.6 | 8.1 | 8.6 | 7.7 | | | | |
| New rage | | 60 | 8.9 | 8.4 | 8.8 | 7.6 | | | | |
| Biostimulant application = 1.5 l ha ⁻¹ (C) | | 90 | 8.6 | 8.1 | 8.7 | 7.7 | | | | |
| 0 8.9 7.9 8.8 7.7 30 8.5 8.0 8.9 7.2 60 8.9 8.0 9.0 7.5 90 8.8 8.0 9.0 7.5 90 8.8 8.0 8.6 7.2 Biostimulant application – 3.0 l ha ⁻¹ (C) 0 8.6 8.2 8.4 7.2 30 8.8 7.8 8.3 7.3 60 8.9 7.9 8.6 7.6 90 8.5 8.0 8.2 7.6 Average 8.7 8.0 8.4 7.4 30 8.6 8.0 8.7 7.4 30 8.6 8.0 8.6 7.4 30 8.6 8.0 8.6 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) 8.6 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) 8.6 | Av | erage | 8.7 | 8.1 | 8.7 | 7.6 | | | | |
| 30 8.5 8.0 8.9 7.2 60 8.9 8.0 9.0 7.5 90 8.8 8.0 8.6 7.2 Average 8.8 8.0 8.8 7.4 Biostimulant application − 3.0 1 ha ⁻¹ (C) 0 8.6 8.2 8.4 7.2 30 8.8 7.8 8.3 7.3 60 8.9 7.9 8.6 7.6 90 8.5 8.0 8.2 7.6 Average 8.7 8.0 8.2 7.6 Average 8.7 8.0 8.7 7.4 30 8.8 8.0 8.7 7.4 Nerage 8.7 8.0 8.7 7.4 Average 8.7 8.0 8.7 7.4 Nerage 8.8 8.0 8.7 7.4 Nerage 8.8 8.0 8.7 7.4 Average 8.7 8.0 8.6 7.6 90 8.8 8.0 8.7 7.4 NIR _{0.05} (Tukey's test) Darkening after 10 min A − 0.12 B − 0.31 C − 0.10 B/A − ns A/B − ns B/C − ns A/C − ns C/B − 0.20 B/C − 0.35 Darkening after 4 h A − 0.42 B − ns C − ns B/A − ns A/B − ns B/C − ns | | В | iostimulant appl | lication – 1.5 l h | na ⁻¹ (C) | | | | | |
| 60 8.9 8.0 9.0 7.5 90 8.8 8.0 8.6 7.2 Average 8.8 8.0 8.6 7.4 Biostimulant application − 3.0 l ha¹ (C) 0 8.6 8.2 8.4 7.2 30 8.8 7.8 8.3 7.3 60 8.9 7.9 8.6 7.6 90 8.5 8.0 8.2 7.6 Average 8.7 8.0 8.4 7.4 Average 8.7 8.0 8.4 7.4 Average 8.8 8.0 8.7 7.4 30 8.6 8.0 8.7 7.4 30 8.6 8.0 8.6 7.6 Average 8.7 8.0 8.7 7.4 30 8.6 8.0 8.6 7.4 10 8.9 8.1 8.8 7.6 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A − 0.12 B − 0.31 C − 0.10 B/A − ns A/B − ns B/C − ns A/C − ns C/B − 0.20 B/C − 0.35 Darkening after 4 h A − 0.42 B − ns C − ns B/A − ns A/B − ns B/C − ns | | 0 | 8.9 | 7.9 | 8.8 | 7.7 | | | | |
| Second | | 30 | 8.5 | 8.0 | 8.9 | 7.2 | | | | |
| Average 8.8 8.0 8.8 7.4 Biostimulant application - 3.0 l ha ⁻¹ (C) 0 8.6 8.2 8.4 7.2 30 8.8 7.8 8.3 7.3 60 8.9 7.9 8.6 7.6 90 8.5 8.0 8.2 7.6 Average 0 8.8 8.0 8.7 7.4 30 8.6 8.0 8.6 7.4 60 8.9 8.1 8.8 7.6 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A - 0.12 B - 0.31 C - 0.10 B/A - ns A/B - ns B/C - ns A/C - ns C/B - 0.20 B/C - 0.35 Darkening after 4 A A A A - 0.42 B - ns C - ns B/A - ns A/B - ns B/C - ns | | 60 | 8.9 | 8.0 | 9.0 | 7.5 | | | | |
| Biostimulant application — 3.0 l ha ⁻¹ (C) 0 8.6 8.2 8.4 7.2 30 8.8 7.8 8.3 7.3 60 8.9 7.9 8.6 7.6 90 8.5 8.0 8.2 7.6 Average 8.7 8.0 8.4 7.4 | | 90 | 8.8 | 8.0 | 8.6 | 7.2 | | | | |
| 0 8.6 8.2 8.4 7.2 30 8.8 7.8 8.3 7.3 60 8.9 7.9 8.6 7.6 90 8.5 8.0 8.2 7.6 Average 0 8.8 8.0 8.7 7.4 30 8.6 8.0 8.6 7.4 60 8.9 8.1 8.8 7.6 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A - 0.12 B - 0.31 C - 0.10 B/A - ns A/B - ns B/C - ns A/C - ns C/B - 0.20 B/C - 0.35 Darkening after 4 h A - 0.42 B - ns C - ns B/A - ns A/B - ns B/C - ns | Av | erage | 8.8 | 8.0 | 8.8 | 7.4 | | | | |
| 30 8.8 7.8 8.3 7.3 60 8.9 7.9 8.6 7.6 90 8.5 8.0 8.2 7.6 Average 0 8.8 8.0 8.7 7.4 30 8.6 8.0 8.6 7.4 60 8.9 8.1 8.8 7.6 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A - 0.12 B - 0.31 C - 0.10 B/A - ns A/B - ns B/C - ns A/C - ns C/B - 0.20 B/C - 0.35 Darkening after 4 h A - 0.42 B - ns C - ns B/A - ns A/B - ns B/C - ns | | В | iostimulant appl | ication - 3.0 1 h | na ⁻¹ (C) | | | | | |
| 60 8.9 7.9 8.6 7.6 90 8.5 8.0 8.2 7.6 Average Average 0 8.8 8.0 8.7 7.4 30 8.6 8.0 8.6 7.4 60 8.9 8.1 8.8 7.6 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A - 0.12 B - 0.31 C - 0.10 B/A - ns A/B - ns B/C - ns A/C - ns C/B - 0.20 B/C - 0.35 Darkening after 4 h A - 0.42 B - ns C - ns B/A - ns A/B - ns B/C - ns | | 0 | 8.6 | 8.2 | 8.4 | 7.2 | | | | |
| 90 8.5 8.0 8.2 7.6 Average 8.7 8.0 8.4 7.4 Average 0 8.8 8.0 8.7 7.4 30 8.6 8.0 8.6 7.4 60 8.9 8.1 8.8 7.6 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A - 0.12 B - 0.31 C - 0.10 B/A - ns A/B - ns B/C - ns A/C - ns C/B - 0.20 B/C - 0.35 Darkening after 4 h A - 0.42 B - ns C - ns B/A - ns A/B - ns B/C - ns B/A - ns A/B - ns B/C - ns | | 30 | 8.8 | 7.8 | 8.3 | 7.3 | | | | |
| Average 8.7 8.0 8.4 7.4 Average 0 8.8 8.0 8.7 7.4 30 8.6 8.0 8.6 7.4 60 8.9 8.1 8.8 7.6 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A - 0.12 B - 0.31 C - 0.10 B/A - ns A/B - ns B/C - ns A/C - ns C/B - 0.20 B/C - 0.35 Darkening after 4 h A - 0.42 B - ns C - ns B/A - ns A/B - ns B/C - ns | | 60 | 8.9 | 7.9 | 8.6 | 7.6 | | | | |
| Average 0 8.8 8.0 8.7 7.4 30 8.6 8.0 8.6 7.4 60 8.9 8.1 8.8 7.6 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A - 0.12 B - 0.31 C - 0.10 B/A - ns A/B - ns B/C - ns A/C - ns C/B - 0.20 B/C - 0.35 Darkening after 4 h A - 0.42 B - ns C - ns B/A - ns A/B - ns B/C - ns | | 90 | 8.5 | 8.0 | 8.2 | 7.6 | | | | |
| 0 8.8 8.0 8.7 7.4 30 8.6 8.0 8.6 7.4 60 8.9 8.1 8.8 7.6 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A - 0.12 B - 0.31 C - 0.10 B/A - ns A/B - ns B/C - ns A/C - ns C/B - 0.20 B/C - 0.35 Darkening after 4 h A - 0.42 B - ns C - ns B/A - ns A/B - ns B/C - ns | Av | erage | 8.7 | 8.0 | 8.4 | 7.4 | | | | |
| 30 8.6 8.0 8.6 7.4 60 8.9 8.1 8.8 7.6 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A - 0.12 B - 0.31 C - 0.10 B/A - ns A/B - ns B/C - ns A/C - ns C/B - 0.20 B/C - 0.35 Darkening after 4 h A - 0.42 B - ns C - ns B/A - ns A/B - ns B/C - ns | | | A | verage | | | | | | |
| 60 8.9 8.1 8.8 7.6 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A - 0.12 B - 0.31 C - 0.10 B/A - ns A/B - ns B/C - ns A/C - ns C/B - 0.20 B/C - 0.35 Darkening after 4 h A - 0.42 B - ns C - ns B/A - ns A/B - ns B/C - ns | | 0 | 8.8 | 8.0 | 8.7 | 7.4 | | | | |
| 90 8.6 8.0 8.5 7.5 Average 8.7 8.0 8.6 7.5 NIR _{0.05} (Tukey's test) Darkening after 10 min A - 0.12 B - 0.31 C - 0.10 B/A - ns A/B - ns B/C - ns A/C - ns C/B - 0.20 B/C - 0.35 Darkening after 4 h A - 0.42 B - ns C - ns B/A - ns A/B - ns B/C - ns | | 30 | 8.6 | 8.0 | 8.6 | 7.4 | | | | |
| | | 60 | 8.9 | 8.1 | 8.8 | 7.6 | | | | |
| $\begin{array}{llllllllllllllllllllllllllllllllllll$ | | 90 | 8.6 | 8.0 | 8.5 | 7.5 | | | | |
| $\begin{array}{llllllllllllllllllllllllllllllllllll$ | | | 8.7 | 8.0 | 8.6 | 7.5 | | | | |
| $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | NIR _{0.05} (Tuk | ey's test) | | | | | | | | |
| $\begin{array}{cccccccccccccccccccccccccccccccccccc$ | Darkening a | | | | | | | | | |
| $A/C - ns \qquad C/B - 0.20 \qquad B/C - 0.35$ $Darkening after 4 h$ $A - 0.42 \qquad B - ns \qquad C - ns$ $B/A - ns \qquad A/B - ns \qquad B/C - ns$ | A – 0.12 | | C - 0.10 | | | | | | | |
| Darkening after 4 h $A-0.42 \qquad B-ns \qquad C-ns$ $B/A-ns \qquad A/B-ns \qquad B/C-ns$ | B/A – ns | A/B - ns | B/C - ns | | | | | | | |
| A - 0.42 $B - ns$ $C - nsB/A - ns$ $A/B - ns$ $B/C - ns$ | A/C - ns $C/B - 0.20$ | | B/C - 0.35 | | | | | | | |
| B/A - ns $A/B - ns$ $B/C - ns$ | Darkening a | after 4 h | | | | | | | | |
| | A - 0.42 | | C-ns | | | | | | | |
| A/C - ns $C/B - ns$ $B/C - ns$ | B/A – ns | A/B - ns | B/C-ns | | | | | | | |
| | A/C – ns | C/B – ns | B/C – ns | | | | | | | |

ns-non-significant

darkening processes in tubers (Keutgen et al. 2014). According to Gugala and Zarzecka (2007), enzymatic darkening of flesh depends on edaphic conditions. Color, on the other hand, depends not only on edaphic conditions but also on agrotechnical conditions.

Statistical analysis showed a significant effect of magnesium fertilization and the application of the biostimulant on the darkening of the flesh of cooked potato tubers after 10 min and 24 h (Table 7). The only exception is the effect of magnesium fertilization on the degree of darkening of cooked flesh after 24 h, as no significant effect was shown. The degree of darkening of the flesh of cooked potato tubers on objects fertilized with magnesium at a dose of 30 kg ha and after the application of the biostimulant at 1.5 l ha was the brightest. On the other hand, the highest intensity of gray coloration of cooked flesh was recorded in tubers harvested from the control object, both after 10 min and after 24 h. The above correlation is not confirmed by Ceglarek et al. (1998), who showed that potatoes grown in soil treated with mineral fertilizers were characterized by a higher intensity of gray color than potatoes in soil supplied manure or green manures. The intensity of darkening depends on the content of phenolic compounds and is largely a genetic trait. The amount of these compounds depends on edaphic conditions, including soil type, meteorological conditions of the growing season, fertilization, the degree of tuber maturity, etc. (Ding et al. 2002). The darkening of the cooked flesh of potato tubers was also differentiated by the timing of the tests. Potatoes evaluated immediately after harvesting were characterized by a lower degree of darkening of the cooked flesh after 10 minutes and 24 hours than tubers evaluated after long-term storage. According to many authors (Keutgen et al. 2014, Wszelaczyńska et al. 2016), long-term storage always deteriorates the quality of tubers in terms of their tendency to flesh darkening. This is often caused by the inability to maintain stable conditions during storage (Pobereżny, Wszelaczyńska 2011, Wszelaczyńska, Poberezny 2011). This research confirmed the negative effect of long-term storage on the tendency for the flesh of tubers to darken. As a result of storage, there was an increase in the darkening of both raw and cooked tuber flesh. Increased propensity for enzymatic and non-enzymatic darkening of flesh under the influence of storage was also reported by Wszelaczyńska (2004), Keutgen et al. (2014) and Wszelaczyńska et al. (2017). The authors conducted a study on other edible varieties, which, same as in our experiment, showed an increase in the propensity of the flesh for darkening. It should be noted, however, that the change in the consumption value of the parameter was small, as the varieties tested in the above studies still belonged to the group of potatoes with a very low propensity to darkening.

TGA content

Regardless of the factors used, the TGA content of the tubers of the Gala variety was at low, both after harvest 35.1 and after storage 37.9 mg kg⁻¹ fresh weight (Table 8). Gugała et al. (2016), in their research on three edible

Table 7 Evaluation of darkening after 10 min and 24 h of cooked potato tubers of the Gala variety depending on the cultivation technology and evaluation date (average of 2015-2017)

| MgO | P | otato tuber eva | aluation date (A |) |
|------------------------------------|----------------------------|-------------------|----------------------|------------|
| fertilization doses (B) | directly aft | er harvest | after s | torage |
| (kg ha ⁻¹) | after 10 min | after 24 h | after 10 min | after 24 h |
| Wit | thout biostimulan | t application, co | ntrol (C) | |
| 0 | 8.7 | 8.4 | 8.6 | 8.3 |
| 30 | 8.8 | 8.6 | 8.7 | 8.4 |
| 60 | 8.7 | 8.3 | 8.4 | 8.2 |
| 90 | 8.6 | 8.3 | 8.3 | 8.1 |
| Average | 8.7 | 8.4 | 8.5 | 8.2 |
| В | iostimulant appl | ication – 1.5 l ł | na ⁻¹ (C) | |
| 0 | 8.9 | 8.6 | 8.9 | 8.5 |
| 30 | 9.0 | 9.0 | 9.0 | 8.5 |
| 60 | 8.8 | 8.5 | 8.3 | 8.3 |
| 90 | 8.8 | 8.4 | 8.5 | 8.3 |
| Average | 8.9 | 8.6 | 8.7 | 8.4 |
| B | iostimulant appl | ication – 3.0 l ł | na-1 (C) | |
| 0 | 8.5 | 8.1 | 8.3 | 8.0 |
| 30 | 8.6 | 8.3 | 8.5 | 8.1 |
| 60 | 8.6 | 8.0 | 8.5 | 7.7 |
| 90 | 8.5 | 8.1 | 8.1 | 8.0 |
| Average | 8.6 | 8.1 | 8.3 | 7.9 |
| | A | verage | | |
| 0 | 8.7 | 8.4 | 8.6 | 8.3 |
| 30 | 8.8 | 8.6 | 8.7 | 8.3 |
| 60 | 8.7 | 8.3 | 8.4 | 8.0 |
| 90 | 8.6 | 8.3 | 8.3 | 8.1 |
| Average | 8.7 | 8.4 | 8.5 | 8.2 |
| NIR _{0.05} (Tukey's test) | | | | |
| Darkening after 10 min | | | | |
| A - 0.11 B - 0.23 | C - 0.13 | | | |
| B/A - ns $A/B - ns$ | $\mathrm{B/C}-\mathrm{ns}$ | | | |
| A/C - ns $C/B - ns$ | B/C-ns | | | |
| Darkening after 24 h | | | | |
| A - 0.08 $B - ns$ | C - 0.13 | | | |
| B/A - ns $A/B - ns$ | $\mathrm{B/C}-\mathrm{ns}$ | | | |
| A/C - ns $C/B - ns$ | B/C-ns | | | |

ns-non-significant

 $\label{thm:control} Table~8$ Total glycoalkaloid content in potato tubers of the Gala variety depending on the cultivation technology and evaluation date (mg kg 1 f. m.), average of 2015-2017

| | MgO | Potato tuber evalu | uation date (A) | |
|---|------------|--|-----------------|--|
| fertilization doses B (kg ha ⁻¹) | | directly after harvest | after storage | |
| | With | out biostimulant application, contro | ol (C) | |
| | 0 | 30,9 | 33,6 | |
| | 30 | 33,2 | 36,6 | |
| | 60 | 34,3 | 38,1 | |
| | 90 | 35,5 | 40,0 | |
| A | verage | 33,5 | 37,1 | |
| | Bio | stimulant application – 1.5 l ha ⁻¹ | (C) | |
| | 0 | 32,9 | 35,6 | |
| | 30 | 34,3 | 36,9 | |
| | 60 | 35,2 | 37,6 | |
| | 90 | 36,6 | 38,4 | |
| Average | | 34,8 | 37,1 | |
| | Bio | stimulant application – 3.0 l ha ⁻¹ | (C) | |
| | 0 | 35,3 | 37,4 | |
| | 30 | 36,2 | 39,1 | |
| | 60 | 37,5 | 40,2 | |
| | 90 | 38,7 | 41,1 | |
| A | verage | 37,0 | 39,5 | |
| | | Average | | |
| | 0 | 33,0 | 35,5 | |
| 30 | | 34,6 | 37,5 | |
| 60 | | 35,7 | 38,6 | |
| 90 | | 37,0 | 39,9 | |
| Average | | 35,1 | 37,9 | |
| VIR _{0.05} (Tuke | ey's test) | | | |
| A - 2.55 | B - 1.27 | C - 0.77 | | |
| B/A - ns | A/B - ns | B/C - ns | | |
| VC – ns | C/B - ns | B/C - ns | | |

ns-non-significant

potato varieties, obtained higher TGA content. The TGA content averaged 91.14 mg kg⁻¹ fresh weight of the Honorata variety tubers up to 92.31 for the Bartek variety tubers. Many authors (Friedman 2006, Wroniak 2006, Zarzyńska 2013, Wszelaczyńska 2020) indicate that the average TGA content

of tubers for consumption is between 20-100 mg kg⁻¹ fresh weight. According to Sengül et al. (2004) and Zarzyńska (2013), edible potato tubers should have a TGA content of less than 50 mg kg⁻¹ FW. In our study, the application of magnesium fertilization as well as that of the biostimulant increased the TGA content in tubers after harvest. Each dose of magnesium increased the TGA content, and the highest content was in tubers from the object fertilized with 90 kg ha⁻¹. Wszelaczyńska et al. (2020) applied magnesium in doses ranging from 20 to 100 kg ha⁻¹ and showed that each increase in the Mg dose raised the TGA content of tubers after harvest. They obtained the highest TGA content following the application of using 100 kg. The supply of magnesium during the growing season of plants can accelerate the intensity of photosynthesis, as a result of which the content of nitrogen compounds in tubers increases (Cakmak, Kirkby 2008, Wszelaczyńska 2020). A similar effect was obtained after the biostimulant application, as the highest TGA content was observed in tubers of plants treated with the highest dose of the biostimulant, i.e. 3.0 l ha⁻¹, during the growing season. Gugała et al. (2016), in an experiment with biostimulants, also obtained a significant increase in the content of TGA in tubers. However, it should be noted that the highest increase in TGA was on the plots where only herbicides were applied, without the addition of biostimulants. According to Bejarano et al. (2000), the TGA content in tubers increases due to stress conditions during the growing season so biostimulants that increase plant resistance to stress conditions may contribute to lower TGA (Du Jardina 2015, Hara 2019). Long-term storage (6 months) resulted in an average 7.5% increase in the TGA content in the tubers studied. Similar results were obtained by Zgórska, Sowa-Niedziałkowska (2005) and by Wszelaczyńska et al. (2020) in their studies, where the TGA content increased significantly after storage. Wszelaczyńska et al. (2020) obtained a significant increase in TGA both after three months (by 22%) and after six months (by 38%). In contrast, in the Zgórska, Sowa-Niedziałkowska (2005) study, a significant increase was found only after a longer storage period of as many as eight months (an increase of 35% at a storage temp. of 4°C and 70% at 8°C).

CONCLUSION

The study showed that the application of magnesium during the potato growing season caused a change in the intensity of tuber flesh color to a more yellow color, and increased the content of TGA. The greatest effect in this regard was produced by the dose of 90 kg MgO ha⁻¹. In addition, the application of magnesium reduced the darkening processes of both raw and cooked tubers. The best results in this regard were obtained after the application of magnesium in soil at 30 and 60 kg ha⁻¹. The application of the biostimulant had a positive effect on the distinguishing features of the con-

sumption value: taste and smell. The best taste and neutral smell were obtained by potato tubers from objects where the biostimulant was applied at a dose of 1.5 l ha⁻¹. Application of the biostimulant increased the TGA content in tubers. Long-term storage generally deteriorated the quality of potatoes of the Gala variety in terms of all tested organoleptic characteristics and increased the TGA content.

REFERENCES

- Alamar M.C., Tosetti R, Landahl S., Bermejo A., Terry L.A. 2017. Assuring potato tuber quality during storage: a future perspective. Front. Plant Sci., 8: 2034. DOI: 10.3389/fpls.2017.02034
- Bejarano L., Mignolet E., Devaux A., Espinola N., Carrasco E., Larondelle Y. 2000. Glycoalkaloids in potato tubers: The effect of variety and drought stress on the a-solanine and a-chaconine contents of potatoes. J. Sci. Food Agric., 80: 2096-2100. DOI: org/10.1002/1097-0010(200011)80:14<2096::AID-JSFA757>3.0.CO;2-6-
- Bergers W.W.A. 1980. A rapid quantitative assay for solanidine glycoalkaloids in potatoes and industrial potato protein. Potato Res. 23: 105-110. DOI: org/10.1007/BF02364030
- Cakmak I., Kirkby E. 2008. Role of magnesium in carbon partitioning and alleviating photo-oxidative damage. Physiol. Plant., 133: 692-704. DOI: org/10.1111/j.1399-3054.2007.01042.x
- Ceglarek F., Płaza A., Buraczyńska D., Jabłońska-Ceglarek R. 1998. Alternative organic fertilization of edible potato in the middle-eastern macro-region. Part I. Fertilizer value of intercrops depending on their use against the background of manure and straw fertilization. Rocz. Nauk Rol. Ser. A, 113(3/4): 173-188. (in Polish) http://yadda.icm.edu.pl/yadda//element/bwmeta1.element.agro-article-e8113f55-4566-480f-9477-c9ecd3af6065
- Chung J.C., Chou S.S., Hwang D.F. 2004. Changes in nitrate and nitrite content of four vegetables during storage at refrigerated and ambient temperatures. Food Addit. Contam., 21: 317-322. DOI: org/10.1080/02652030410001668763
- Ciećko Z., Rogozińska I., Żółnowski A., Wyszkowski A. 2005. The influence of potassium fertilization, at different N and P levels, on the culinary features of potato tubers. Biul. IHAR, 237/238:151-159. (in Polish) https://www.researchgate.net/publication/249340541
- Devaux A., Kromann P., Ortiz O. 2014. Potatoes for sustainable global food security. Potato Res., 57: 185-199. DOI: org/10.1007/s11540-014-9265-1
- Ding Ch.-K., Ueda Y., Wang Ch. 2002. Inhibition of loquart enzymatic browning by sulfhydryl compounds. Food Chem., 76: 213-218. DOI: org/10.1016/S0308-8146(01)00270-9
- Du Jardin P. 2015. Plant biostimulants: definition, concept, main categories and regulation. Sci. Hortic., 196: 3-14. DOI: org/10.1016/j.scienta.2015.09.021
- Flis B., Zimnoch-Guzowska E., Mańkowski D. 2012. Correlations among yield, taste, tuber characteristics and mineral contents of potato cultivars grown at different growing conditions. J. Agric. Sci., 4(7): 197-207. DOI: 10.5539/jas.v4n7p197
- Friedman M. 2006. Potato glycoalkaloids and metabolites: Roles in the plant and in the diet. J. Agric. Food Chem., 54: 8655-8681. DOI: org/10.1021/jf061471t
- Ginzberg I., Tokuhisa J.G., Veilleux R.E. 2009. Potato steroidal glycoalkaloids: Biosynthesis and genetic manipulation. Potato Res, 52: 1-15. DOI: org/10.1007/s11540-008-9103-4
- Grudzińska M., Czerko Z., Wierzbicka A., Borowska-Komenda M. 2016. Changes in the content of reducing sugars and sucrose in tubers of 11 potato cultivars during long term storage at 5 and 8°C. Acta Agrophys, 23(1): 31-38. (in Polish) http://www.acta-agrophysica.org//Changes-in-the-content-of-reducing-sugars-and-sucrose-in-tubers-of-11-potato-cultivars, 104978,0,2.html
- Gugała M., Zarzecka K. 2007. Effect of soil tillage systems and herbicides on consumption value

- of potato tubers of Wiking cultivar. Acta Sci. Pol., 6(2): 29-37. (in Polish) https://biblioteka.nauki.pl/articles/46864
- Gugała M., Zarzecka K., Dołęga H., Niewęgłowski M., Sikorska A. 2016. The effect of biostimulants and herbicides on glycoalkaloid accumulation in potato. Plant Soil Environ, 62(6): 256-260. DOI: 10.17221/187/2016-PSE
- Haase N.U. 2010. Glycoalkaloid concentration in potato tubers related to storage and consumer offer. Potato Res., 53: 297-307. DOI: org/10.1007/s11540-010-9162-1
- Hamouz K., Lachman J., Dvořák P., Jůzl M., Pivec V. 2006. The effect of site conditions, variety and fertilization on the content of polyphenols in potato tubers. Plant. Soil Environ., 52(9): 407-412. DOI: org/10.17221/3459-PSE
- Hara P. 2019. The role of biostimulators in potato cultivation. Pol. Potato, 29(2): 18-24. (in Polish) https://agro.icm.edu.pl/agro/element/bwmeta1.element.agro-e9cca208-75f0-4b2c-a277-1867a617240f
- Keutgen A.J., Pobereżny J., Wszelaczyńska E., Murawska B., Spychaj-Fabisiak E. 2014. Influence of storage on darkening processes in potato tubers (Solanum tuberosum L.) and their health properties. Inż. Apar. Chem., 53(2): 086-088. (in Polish) http://yadda.icm.edu.pl//baztech/element/bwmeta1.element.baztech-01d7da64-ab6a-42ee-8919-9a050fa62cd4?q =210c05d0-a832-4c66-aa88-20c26559d66a\$4&qt=IN_PAGE
- Kidoń M., Czapski J. 2010. Colour changes of anthocyanin pigments from purple carrot juice in aqueous solutions at various pH values. Acta Agroph., 15(2): 333-345. (in Polish) http://www.acta-agrophysica.org/Colour-changes-of-anthocyanin-pigments-from-purple-carrot-juice-in-aqueous-solutions,107302,0,2.html
- Krochmal-Marczak B., Sawicka B., Kiełtyka-Dadasiewicz A., Bienia B. 2016. Influence of storage and climatic conditions on the quality tuber flesh potato cultivated in the organic system. Fragm. Agron., 33(2): 44-54. (in Polish) https://www.researchgate.net/publication/308473680
- Krzysztofik B., Skonieczny P. 2010. Impact of the period of storage on changes of physical properties of potato tubers. Inż. Rol., 4(122): 135-141. (in Polish) http://yadda.icm.edu.pl/baztech//element/bwmeta1.element.baztech-article-BAR0-0056-0058?q=bwmeta1.element.baztech-volume-1429-7264-inzynieria_rolnicza-2010- r__14_nr_4;17&qt=CHILDREN-STATELESS
- Leszczyński W. 2000. *Quality of consumer potato*. Żywność, Supl., 4(25): 5-27. (in Polish) http:///journal.pttz.org/wp-content/uploads/2018/01/01_Leszczynski.pdf
- Lisińska G. 2006. Technological value and consumption quality of Polish potato varieties. Zesz. Probl. Post. Nauk Roln., 511: 81-94. (in Polish) http://agro.icm.edu.pl/agro/element//bwmeta1.element.agro-article-636f715c-87a5-4d3d-a238-8a8a5b4a44ca
- Machado R.M.D., Toledo M.C.F., Garcia L.C. 2007. Effect of light and temperature on the formation of glycoalkaloids in potato tubers. Food Control, 18: 503-508. DOI: org/10.1016/j. foodcont.2005.12.008
- Osowski J., Erlichowski T., Urbanowicz J. 2017. The effect of potassium, magnesium and sulphur fertilization on the yield, raw tuber darkening and occurrence of black scurf and early blight of potato tubers. Fragm. Agron., 34(1): 49-59. (in Polish) https://pta.up.poznan.pl//pdf/2017/FA%2034(1)%202017%20Osowski.pdf
- Płaza A. 2010. Consumption value of potato tubers fertilized with intercrops in central-eastern Poland. Zesz. Probl. Post. Nauk Rol., 557: 193-199. (in Polish) https://www.infona.pl//resource/bwmeta1.element.dl-catalog-be6525b4-d8a6-4bf8-828d-7ad3b9d08980
- Płaza A., Ceglarek F., Królikowska M. 2010. The influence of intercrops and farmyard manure fertilization in changeable weather conditions on consumption value of potato tubers. J. Cent. Eur. Agric., 11(1): 47-54. DOI: org/10.5513/JCEA01/11.1.799
- Pobereżny J., Wszelaczyńska E. 2011. Effect of bioelements (N, K, Mg) and long-term storage of potato tubers on quantitative and qualitative losses. Part II. Content of dry matter and starch. J. Elem., 16(2): 237-246. DOI: 10.5601/jelem.2011.16.2.07

- Pyryt B., Kolenda H. 2009. Characteristics of sensory quality of cooked tubers depending on the potato variety and cooking method. Bromatol. Chem. Toksykol., 42(3): 386-390. (in Polish) https://www.ptfarm.pl/wydawnictwa/czasopisma/bromatologia-i-chemiatoksykologiczna//117/-/12627
- Rezaee M., Almasi M., Majdabadi F.A., Minaei S., Khodadadi M. 2011. Potato sprout inhibition and tuber quality after post-harvest treatment with gamma irradiation on different dates. J. Agric. Sci. Tech., 13: 829-842. http://journals.modares.ac.ir/article-23-2714-en.html
- Rogozińska I., Wszelaczyńska E., Pińska M. 2004. Effect of magnesium on consumption and technological quality of cv. Mila potato tubers. J. Elementol., 9(4): 707-716. (in Polish) https://agro.icm.edu.pl/agro/element/bwmeta1.element.dl-catalog-74ddd819-753f-4098-927f-4b506f046f4e
- Roztropowicz S., Czerko Z., Głuska A., Goliszewski W., Gruczek T., Lis B., Lutomirska B., Nowacki W., Wierzejska-Bujakowska A., Zarzyńska K., Zgórska K. 1999. Methodology of observation, measurement and sampling in agrotechnical potato experiments. Wyd. IHAR, Jadwisin, 1-50.
- Rytel E., Lisińska G., Kozicka-Pytlarz M. 2008. The effect of cultivation methods on the quality of potato. Zesz. Probl. Post. Nauk Rol., 530: 259-269. (in Polish) http://www.zppnr.sggw. pl/530-26.pdf
- Rytel E. 2012. Changes in the levels of glycoalkaloids and nitrates after the dehydration of cooked potatoes. Am. J. Potato Res, 89: 501-507. DOI: org/10.1007/s12230-012-9273-0
- Sawicka B., Barbaś P., Bienia B., Krochmal-Marczak B., Wójcik S. 2016. Research quality French fries with tubers of early potato cultivars under different nitrogen fertilization conditions. TPJ, 3(48): 115-125. (in Polish) https://www.researchgate.net/publication/303693531
- Sengül M., Keles F., Keles M.S. 2004. The effect of storage conditions (temperature, light, time) and variety on the glycoalkaloid content of potato tubers and sprouts. Food Control, 15: 281-286. DOI: org/10.1016/S0956-7135(03)00077-X
- Styszko L., Kamasa J. 2007. Relationship between resistance of potato cultivars to pathogens and taste of tubers in the years of different level yielding. Prog. Plant Prot., 47(2): 343-347. (in Polish) https://agro.icm.edu.pl/agro/element/bwmeta1.element.agro-article-a0a0e16a-4c6c-4a84-aa21-0a308a0e1e24
- Tajner-Czopek A., Jarych-Szyszka M., Lisińska G. 2008. Changes in glycoalkaloids content of potatoes destined for consumption. Food Chem., 106: 706-711. DOI: org/10.1016/ /j.foodchem.2007.06.034
- Tilahun S., An H.S., Hwang I.G., Choi J.H., Baek M.W., Choi H. R., Park D.S., Jeong C.S. 2020. Prediction of α -solanine and α -chaconine in potato tubers from hunter color values and VIS/NIR spectra. J. Food Qual. 2020. DOI: org/10.1155/2020/8884219
- Wroniak J. 2006. Nutritional values of the edible potato. Pol. Potato, 16(2): 17-20. (in Polish) https://agro.icm.edu.pl/agro/element/bwmeta1.element.agro-article-44f29928-35e4-4381-b32e-a861ed15115c
- Wszelaczyńska E. 2004. Effect of magnesium fertilization on organic acid content and flesh darkening of potato tubers of Mila variety. Acta Sci. Pol., Agricultura, 3(1): 175-1863. (in Polish) https://agro.icm.edu.pl/agro/element/bwmeta1.element.agro-article-90799410-fb8f-4214-8075-2577f1b1da93
- Wszelaczyńska E., Pobereżny J. 2011. Effect of bioelements (N, K, Mg) and long-term storage of potato tubers on quantitative and qualitative losses. Part 1. Natural losses. J. Elem. 16(1): 135-142. DOI: 10.5601/jelem.2011.16.1.135-142
- Wszelaczyńska E., Pobereżny J., Żary-Sikorska E., Mareček J. 2013. Influence of growing location and tuber storage on selected culinary traits of three potato varieties. Inż. Apar. Chem., 52(2): 68-70. (in Polish) https://yadda.icm.edu.pl/baztech/element/bwmeta1.element.baztech-article-BPP4-0003-0022

- Wszelaczyńska E., Pobereżny J. Wichrowska D. 2016. Yielding and size of losses after storage of potato (Solanum tuberosum L.) cultivar 'Satina' depending on the farming system and the soil fertilizer application. Fresen Environ. Bull., 25(8): 3159-3168. https://www.researchgate.net/publication/309317755
- Wszelaczyńska E., Pobereżny J., Gościnna K., Chmielewski J., Łaba S. 2017. The darkening of potato tubers opportunities to reduce it. Przem. Spoż., 71(11): 31-34. (in Polish) DOI: 10.15199/65.2017.11.6
- Wszelaczyńska E., Pobereżny J., Kozera W., Knapowski T., Pawelzik E. Spychaj-Fabisiak E. 2020. Effect of magnesium supply and storage time on anti-nutritive compounds in potato tubers. Agronomy 10(3): 339. https://doi.org/10.3390/agronomy10030339
- Zarzecka K., Gugała M., Milewska A. 2010. Effects of new generation insecticides on culinary traits of potatoes. Zesz. Probl. Post. Nauk Rol., 557: 201-208. (in Polish) https://www.infona.pl/resource/bwmeta1.element.dl-catalog-02e2294b-9883-4881-a56f-8d4ca007a06b
- Zarzecka K. Gugała M. 2011. Selected qualitative characteristics of edible potato tubers from the Podlasie Region. Bromatol. Chem. Toksykol., 44(1): 38-42. (in Polish) http://agro.icm. edu.pl/agro/element/bwmeta1.element.dl-catalog-8f214d2b-8ef5-4b6f-9d90-5703ea265367?q =bwmeta1.element.dl-catalog-650b5d6a-3b87-43a1-bd5d-6ecb36bd5a89;5&qt=CHILDREN-STATELESS
- Zarzyńska K., Goliszewski W. 2006. Potato production in organic and integrated system and tuber quality. Pam. Puł., 142: 617-626. (in Polish) https://agro.icm.edu.pl/agro/element/bwmeta1. element.dl-catalog-4bde4a87-d23e-43a0-a117-05a968497ca7
- Zarzyńska K., Wroniak J. 2007. Differences in quality of potato tubers growing in organic system depending on some agronomical factors. J. Agric. Eng. Res., 52(4): 108-114. (in Polish) https://bibliotekanauki.pl/articles/337683
- Zarzyńska K. 2013. Chemical composition of potato tubers in relation to crop production system and environmental conditions. J. Agric. Sci. Technol., B 3: 689-695.
- Zgórska K., Sowa-Niedziałkowska G. 2005. The influence of storage temperature and cultivar on quality changes in potato tubers during long term storage. Pam. Puł., 139: 327-336. https://agro.icm.edu.pl/agro/element/bwmeta1.element.dl-catalog-132f7f06-1c44-4d66-9d7a-8fdef2b379f5
- Zgórska K., Czerko Z., Grudzińska M. 2006. Influence of storage conditions on some culinary and technological characteristics of tubers of selected potato varieties. Zesz. Probl. Post. Nauk Rol., 511: 567-578. (in Polish) https://www.infona.pl/resource/bwmeta1.element.agro-article-9a1a6f71-4783-4b02-93eb-5e5131691bd0
- Zgórska K., Grudzińska M. 2012. Changes in selected quality parameters of potato tubers during storage. Acta Agroph, 19(1): 203-214. (in Polish) http://www.acta-agrophysica.org/,104771,0,2.html. Zgórska K. 2013. Use of potatoes for food and industrial purposes. Pol. J. Food Eng., 3(7): 5-9. (in Polish) http://www.acta-agrophysica.org/,104771,0,2.html
- Zimnoch-Guzowska E., Flis B. 2006. Genetic basis of potato quality traits. Zesz. Probl. Post. Nauk Rol., 51: 23-36. (in Polish) https://www.infona.pl/resource/bwmeta1.element.agro-article-f07277f2-8e3f-4fa6-bb28-4bc98b2ee8ac